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Outline

Doctoral Thesis

**Study on Acyl-CoA Synthetase Involved
in Fatty Acid Utilization of Yeast
Malassezia spp.**

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THESIS OUTLINE

INTRODUCTION

Malassezia spp. are common habitat yeast in human or homothermic animals' skin. All species of this yeast does not possess fatty acid synthase causing the dependence on exogenous lipids from the host for their growth and proliferation. In immunocompromised individuals, this yeast can cause various skin diseases such as seborrheic dermatitis, pityriasis versicolor, dog's otitis, and dandruff. One of the pathogenicity traits of *Malassezia* spp. is the lipase secretion which degrade host's lipid into fatty acids. Some of the fatty acids can penetrate through the epidermal barrier of the skin causing inflammation. The other fatty acids will be consumed by the yeast for its growth. However, how a fatty acid is utilized by *Malassezia* spp. and how is it related to the pathogenicity remain to be clarified.

1. Identification of acyl-CoA synthetase (ACS) involved in fatty acid utilization in *Malassezia* spp.

Before a fatty acid can enter any of the metabolism pathways, it needs to be activated to fatty acyl-CoA by ACS (fatty acid:CoA ligase, E.C. 6.2.1.3). This enzyme catalyzes the thioesterification of fatty acid and CoA using ATP. Six genes encoding ACS in *Saccharomyces cerevisiae* have been well studied with *ScFAA1* and *ScFAA4* as the main fatty acid activators. Deletion of these two genes resulted in the inability of *S. cerevisiae* to grow in medium containing fatty acids and fatty acid synthase inhibitor cerulenin. In this study, complementation of genes encoding *FAA1* of *M. pachydermatis* (Malapachy_0054, *MpFAA1*), *M. globosa* (MGL_3626, *MgFAA1*), and *M. sympodialis* (MSYG_3835, *MsFAA1*) into *S. cerevisiae* *faa1*Δ *faa4* Δ restored the growth of *S. cerevisiae* mutant in minimal medium containing cerulenin and long-chain fatty acid C12:0, C14:0, C16:0, and C18. Gene expression level analysis by qRT-PCR suggested that *MpFAA1* was induced in the presence of palmitic acid and oleic acid, while *MpFAA2* was induced only in palmitic acid. These results suggested one of the possibilities why *M. pachydermatis* is able to grow more efficiently in the medium lacking of fatty acids, that its *FAA1* and *FAA2* are highly expressed in fatty acids, unlike the other species.

2. Effect of ACS inhibition and its potential as new drug targets for *Malassezia* treatment.

ACSs are assumed to be potential as new drug targets because FAS is absent in all *Malassezia* spp. Inhibition of activation of exogenous fatty acids can disrupt the fatty acid

metabolism in this yeast and probably cause the cell death. Triacsin C, a potent ACS inhibitor, hampered the growth of *M. pachydermatis*, *M. globosa*, and *M. sympodialis* in rich medium mDixon, with *M. pachydermatis* as the most sensitive species among the three. Supplementation of this inhibitor in minimal medium containing cerulenin and fatty acids also block the restore of *MpFAA1*, *MgFAA1*, and *MsFAA1* in *S. cerevisiae faa1Δ faa4 Δ*. However, the *S. cerevisiae* WT could still grow in the presence of Triacsin C, showing that *Malassezia FAA1* is the target of Triacsin C but not that of *S. cerevisiae*. This inhibitor also affected the lipid droplet formation (smaller size) in *Malassezia* and interfere the lipid synthesis for the cell wall formation, causing the weaker cell membrane. Addition of Triacsin C also reduced the acyl-CoA products and increased FFAs were detected. Thus, *Malassezia Faa1p* is indeed potential as a new drug target for *Malassezia* treatment.

3. Purification and Characterization of *Malassezia* ACS protein.

Purification of *Malassezia* ACS proteins, especially *Faa1p*, was carried out to understand more about the characteristics of these enzymes. *MpFAA1* and *MgFAA1* ORF which have no introns were tagged with 6xhis-tag at their N-terminal and successfully cloned into BL21 (DE3) *E. coli* strain for *Faa1* protein purification. The purification also resulted the clear band for the target protein with the correct size. Even though few impurities still appeared, this is the first attempt (to our knowledge) that demonstrate the purification of *Malassezia* proteins using *E. coli*.

Conclusion and Future Perspective

This study showed that three *Malassezia FAA1s* have similar functions with that of *S. cerevisiae* in utilization of fatty acid for membrane lipid synthesis. The ACS inhibitor Triacsin C block the restore of *Malassezia FAA1* in *S. cerevisiae faa1Δ faa4Δ*. This inhibitor also reduced the growth of *M. pachydermatis*, *M. globosa*, and *M. sympodialis* but not totally killed the yeasts. The LD & biofilm formation in the three *Malassezia*, especially *M. pachydermatis* were diminished by Triacsin C. These results suggest that *Faa1p* is potential as a new drug target for *Malassezia* treatment. However, further study is required specifically on *Faa1* protein study. In the last chapter, *M.pachydermatis Faa1p* was successfully extracted and purified from *E.coli* BL21 (DE3) pLyss. The enzyme activity and protein structure change of *Faa1p* is potential to develop a more effective anti-*Malassezia* drug.