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Outline

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Title: Isotopomeric analysis on the production and consumption processes of nitrous oxide accumulated in and emitted from Japanese tea field soil

(日本の茶園土壌において蓄積かつ放出される一酸化二窒素の生成および消滅過程に関するアイソトポマー解析)

Nitrous oxide (N_2O) is a greenhouse gas in the troposphere and a potential destroyer of stratospheric ozone layer. N_2O concentration has risen rapidly from 1750 until now, and it will continue increasing in the future. Natural soils and agricultural soils are the biggest source of atmospheric N_2O . In the soils, N_2O is produced through microbial processes, nitrification and denitrification. The high nitrogenous fertilizer application caused soil acidification and large N_2O emission from Japanese tea field.

The soil N_2O gas, N_2O flux, soil, soil water, and rainwater samples were collected in 2011 and 2012. The concentrations and isotopemer ratios of N_2O and inorganic nitrogen were analyzed to investigate the sources and sinks of N_2O in Japanese tea field under conventional, lime nitrogen, and DCD (dicyandiamide) treatments.

According to the specific ^{15}N site preference (SP) values of bacterial nitrification,

bacterial denitrifier-denitrification, nitrifier-denitrification, and fungal denitrification, it was suggested that bacterial denitrification was the dominant process of N₂O production on most days in two parallel plots (Plot I and Plot II) under conventional treatment, irrespective of N₂O reduction occurring before or after mixing. However, on 31 May 2011 (after one of fertilizations) during which soil temperatures were 15.8°C to 17.9°C and water-filled pore space (WFPS) was 0.73 to 0.89 in the upper layer of Plot I and deeper layers of Plot II, and on 18 October 2012 during which soil temperatures were 19.7°C to 20.1°C and WFPS was 0.61 to 0.74 in Plot II, bacterial denitrification and bacterial nitrification (or fungal denitrification) contributed nearly equal shares on the produced N₂O. Moreover, on 4 October 2012 (35 cm depth; WFPS was 0.73) and 18 October 2012 (10 cm depth; soil temperature was 19.7°C and WFPS was 0.68) in Plot I, bacterial nitrification (or fungal denitrification) contributed more N₂O than bacterial denitrification.

During the formation of N₂O through denitrifier denitrification process, the fractions of O exchange between soil water and N₂O precursors were approximately 80% and 77%, respectively, at 10 cm and 20 cm depth soils. The net ¹⁸O isotope effects for denitrification (NO₃⁻ reduction to N₂O) were estimated as 35‰ and 33‰, respectively, at 10 cm and 20 cm depth soil.

In the tea field, lime nitrogen effectively suppressed N₂O emission by inhibiting nitrification process. The inhibition effect by lime nitrogen (CaCN₂) could last for one month or shorter. The inhibition effect decreased with time. However, DCD treatment enhanced N₂O emission. The proportion of N₂O produced through bacterial nitrification process was ascended by DCD. Ineffective inhibition by DCD could be attributed to repeated application year by year.